Phenolic Constituents of Gnetum klossii

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Received November 15, 2002

Four new phenolic derivatives, gnetofurans A-C (1-3) and dihydropinosylvindiol (4), were isolated from a methanol-soluble extract of the stems of Gnetum klossii, together with nine known compounds [gnetifolin F (5), isorhapontigenin, gnetulin, gnetins E and C, latifolol, gnetol, (-)- ϵ -viniferin, and *trans*-resveratrol]. The structures of the new compounds were determined by spectral data analysis.

The genus Gnetum (Gnetaceae) consists of about 40 species distributed in South America (Amazon region), Southwest Africa, and the tropical and subtropical zones of Asia.1 Various species have been used in folk medicine for the treatment of arthritis, bronchitis, and asthma.² The genus Gnetum is well known for its abundant polyphenolic constituents.^{2,3} In a continuation of our phytochemical studies on *Gnetum* species, ^{4–9} we report herein the isolation and structure elucidation of four new phenolic derivatives, gnetofurans A-C (1-3) and dihydropinosylvindiol (4), along with nine known compounds from a methanol-soluble part of the stems of *G. klossii* Merr. The known constituents were gnetifolin F (5),¹⁰ isorhapontigenin, gnetol, (-)- ϵ viniferin,¹¹ gnetulin,⁴ gnetins C and E,¹² latifolol,⁷ and trans-resveratrol.13



Gnetofuran A (1), a white amorphous powder, showed a positive reaction to Gibbs reagent. The negative FABMS exhibited a $[M - H]^-$ ion peak at m/z 435, and the molecular formula C₂₅H₂₄O₇ was deduced from the HR-FABMS at m/z 435.1449 [M - H]⁻ (calcd 435.1444). The ¹H NMR, ¹H-¹H COSY, and ¹H-¹H long-range COSY spectra exhibited the presence of a set of meta-coupled H atoms in an AB system on a tetrasubstituted benzene ring (ring A) [δ 7.00 (1H, br s, H-6), 6.98 (1H, br s, H-2)], a set



Figure 1. CH long-range correlations (→) of compounds 1 and 2 from their HMBC and of compounds 4 and 5 from their COLOC spectra, HH long-range correlations (-) of 1 and 4 from their ¹H-¹H long-range COSY spectra, and NOE interactions (- - -) of compounds 1, 2, and 5 from their NOESY spectra.

of meta-coupled protons in an A2B system on the 1,3,5trisubstituted benzene ring (ring B) [δ 6.41 (2H, d, J = 2.1Hz, H-10(14), 6.14 (1H, t, J = 2.1 Hz, H-12)], and ABX system protons for a 1,2,4-trisubstituted benzene ring (ring C) [δ 6.92 (1H, d, J = 2.1 Hz, H-2'), 6.75 (1H, dd, J = 2.1, 8.1 Hz, H-6'), 6.67 (1H, d, J = 8.1 Hz, H-5')]. These spectra also showed a pair of *trans*-coupled olefinic H atoms [δ 6.89 (1H, d, J = 16.4 Hz, H-7), 6.78 (1H, d, J = 16.4 Hz, H-8)]and a spin system of four aliphatic protons [δ 5.45 (1H, d, J = 5.4 Hz, H-7'), 3.74 (2H, m, 2H-9'), 3.41 (1H, dt, J =5.4, 12.2 Hz, H-8')]. The ¹H NMR spectrum also showed two signals for methoxyl groups [δ 3.73 (3H, s, OCH₃-3), 3.69 (3H, s, O CH_3 -3')], three for phenolic hydroxyl groups [*b* 8.06 (2H, s, O*H*-11, 13), 7.47 (1H, s, OH-4')], and a signal for an aliphatic hydroxyl group [δ 4.01(1H, br s, OH-9')]. The correlations of H-7/H-2(6), H-8/H-10(14), H-8'/H-6, and H-7'/H-2'(6') in the long-range ¹H-¹H COSY spectrum (Figure 1) and of H-7/C-2(6), H-8/C-10(14), H-8'/C-6, and

10.1021/np0205320 CCC: \$25.00 © 2003 American Chemical Society and American Society of Pharmacognosy Published on Web 03/22/2003

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H-7'/C-2'(6') in the HMBC spectrum revealed the linkages between C-7/C-1, C-8/C-9, C-8'/C-5, and C-7'/C-1', respectively. The two methoxyl groups were assigned to their respective positions by the HMBC correlations of OCH₃-3,H-2/C-3 and OCH3-3',OH-4',H-2'/C-3' and further confirmed by the interactions observed in the NOESY spectrum between OCH₃-3/H-2 and OCH₃-3'/H-2' (Figure 1). The presence of the dihydrofuran ring was deduced by considering the degrees of unsaturation and confirmed by the correlation of H-7' (δ 5.45) with C-4 (δ 149.3) in the HMBC spectrum. In the NOESY spectrum (Figure 1), the interaction between H-7' and H-9' revealed the relative stereochemistry at dihydrofuran to be trans. The chemical shift values of 1 were assigned with the help of HMQC, HMBC, ¹H-¹H COSY, ¹H-¹H long-range COSY, and NOESY spectra (Figure 1). Biogenetically, compound **1** was presumed to be formed by the oxidative coupling of isorhapontigenin (trans-3-methoxy-4,11,13-trihydroxystilbene) with coniferyl alcohol.

Gnetofuran B (2), a white amorphous powder, showed a positive reaction to Gibbs reagent. The EIMS exhibited a $[M]^+$ ion peak at m/z 286. The molecular formula $C_{16}H_{14}O_5$ was deduced from the HREIMS at m/z 286.0834 [M]+ (calcd 286.0841). The characteristic UV absorption bands at 321 and 303 nm showed the presence of a furan ring in the molecule. The ¹H NMR spectrum showed three metacoupled protons [δ 7.00 (1H, t, J = 2.0 Hz, H-6'), 6.99 (1H, t, J = 2.0 Hz, H-2'), 6.44 (1H, t, J = 2.0 Hz, H-4')], two *ortho*-coupled protons [δ 7.10 (1H, dd, J = 8.8, 0.9 Hz, H-7), 6.87 (1H, d, J = 8.8 Hz, H-6)], and one long-range coupled proton [δ 7.35 (1H, d, J = 0.9 Hz, H-3)] in the aromatic region. The signals for two methoxyl [δ 4.05 (3H, s, OCH₃-4), 3.85 (3H, s, OCH₃-3')] and two phenolic hydroxyl [δ 8.45 (1H, s, OH-5'), 7.46 (1H, s, OH-5)] groups were also observed in the ¹H NMR spectrum. The correlations between three *meta*-coupled protons to each other in the ¹H⁻¹H long-range COSY spectrum indicated that they are related to a 1,3,5-trisubstituted benzene ring (ring B). The correlations between H-2'(6')/C-2 and H-3/C-4(3a,7a) in the HMBC spectrum (Figure 1) revealed the linkages between C-1'/C-2, and C-3/C-3a, respectively. All chemical shifts were assigned by the combination of HMQC, HMBC, NOESY, ¹H-¹H COSY, and ¹H-¹H long-range COSY spectra (Figure 1).

Gnetofuran C (**3**), a white amorphous powder, showed a positive reaction to Gibbs reagent. The EIMS exhibited a $[M]^+$ ion peak at m/z 272. The $[M]^+$ ion peak at m/z 272.0677 (calcd 272.0685) in the HREIMS showed the molecular formula to be $C_{15}H_{12}O_5$. Compound **3** showed a general resemblance to **2** except for the replacement of a methoxyl by a hydroxyl group in ring B, due to which the three *meta*-coupled protons appeared as an A₂B system [δ 6.91 (2H, br s, H-2',6'), 6.04 (1H, br s, H-4')] and two phenolic hydroxyl protons appeared as a singlet with double integration [δ 8.43 (2H, s, OH-3',5')]. All chemical shift values were assigned in comparison to compound **2**.

Compounds **2** and **3** are phytoalexins with structural similarity to moracins A and B, and biogenetically these phytoalexins seem to be derived from moracin M.¹⁴

Dihydropinosylvindiol (4), a white amorphous powder, showed a positive reaction to Gibbs reagent. The negative FABMS exhibited a $[M - H]^-$ ion peak at m/z 245, and the molecular formula $C_{14}H_{14}O_4$ was supported by the negative HRFABMS (m/z 245.0808, calcd 245.0814). The ion peak observed at m/z 228 [M - H₂O]⁺ in the EIMS represents the loss of a water molecule. The ¹H NMR spectrum showed five aromatic protons, assignable to a

1-substituted benzene ring [δ 7.28 (2H, br d, J = 8.0 Hz, H-2,6), 7.19 (3H, m, H-3,4,5)], and a set of meta-coupled protons in an A₂B system on a 1,3,5-trisubstituted benzene ring [δ 6.30 (2H, d, J = 2.4 Hz, H-10(14), 6.20 (1H, t, J =2.4 Hz, H-12)]. Two mutually coupled aliphatic methines $[\delta 4.72 (1H, dd, J = 6.0, 4.4 Hz, H-7), 4.63 (1H, dd, J =$ 6.0, 4.4 Hz, H-8)] and two aliphatic hydroxyl protons [δ 4.08 (1H, d, J = 4.4 Hz, OH-7), 4.05 (1H, d, J = 4.4 Hz, OH-8)] together with two phenolic hydroxyl protons [δ 7.97 (2H, s, OH-11,13)] were also observed in this spectrum. The correlations between H-7/H-2(6), H-8/H-10(14) in the longrange ¹H-¹H COSY spectrum (Figure 1) and between C-2(6)/H-7, C-10(14)/H-8 in the COLOC spectrum (Figure 1) revealed the C-7/C-1 and C-8/C-9 linkages, respectively. The two aliphatic methines appeared as clear doubledoublets at δ 4.60 and 4.69 (J = 6.0 Hz) when the ¹H NMR spectrum of 4 was recorded in CD₃OD. Compound 4 (dihydropinosylvindiol) also belongs to phytoalexin and seems to be derived from pinosylvin¹¹ isolated from *Gnetum* parvifolium.

The ¹H and ¹³C NMR spectral data of gnetifolin F (5) and its relative stereochemistry are being reported for the first time in the native state; previously this compound was reported as the acetate form.¹⁰

Experimental Section

General Experimental Procedures. Optical rotations were recorded on a JASCO P-1020 polarimeter, and UV spectra were recorded on a Shimadzu UV 2200 spectrometer. NMR spectra were recorded on a JEOL EX-400 or AL 300 spectrometer with tetramethylsilane (TMS) as an internal reference. EIMS and negative ion FABMS were measured on a JEOL JMS-DX 300 spectrometer equipped with a JMA 3500 data analysis system. Silica gel 60 (70–230 mesh, Merck) and Sephadex LH-20 (Pharmacia) were used for column chromatography. Kieselgel 60 F_{254} (Merck) was used for analytical and preparative TLC.

Plant Material. *Gnetum klossii* Merr. was cultivated at Bogor Botanical Garden, Bogor, Indonesia, from where its stems were collected in April 2001 by one of the co-authors (D.D.), and a voucher specimen number GN-004 was deposited at the herbarium of Gifu Prefectural Institute of Health and Environmental Sciences.

Extraction and Isolation. The dried stems of G. klossii (550 g) were powdered and extracted with MeOH (static, 2 L \times 3 days \times three times). The methanol extract (13 g) was chromatographed on silica gel eluted with a mixture of AcOEt-CHCl₃-MeOH-H₂O (75:75:30:5) to give 10 fractions (each 60 mL, A-J). Compound 2 (9 mg), isorhapontigenin (12 mg), and resveratrol (67 mg) were purified from fraction B after column chromatography over Sephadex LH-20 (MeOH) followed by preparative TLC [CHCl₃-MeOH, 95:5]. Fractions C and D were combined and chromatographed over Sephadex LH-20 eluted with MeOH to give 18 subfractions (each 60 mL, $C-D_{1-18}$). Compound 4 (12 mg) was obtained from fraction C-D₄ by column chromatography over silica gel (CHCl₃-MeOH, 95:5) followed by preparative TLC [benzene-AcOEt-MeOH-H₂O, 20:14:6:1], and gnetifolin F (26 mg) was purified from fraction $C-D_5$ in a fashion similar to that for compound 4. Compounds 1 (11 mg), 3 (7 mg), and gnetulin (51 mg) were purified from fraction C-D₉₋₁₀ by preparative TLC [CHCl₃-MeOH–H₂O, 30:10:1]. Fraction $C-D_{11-12}$ was treated similarly to fraction $C-D_{9-10}$ to afford gnetol (8 mg). Gnetin C (23 mg) and ϵ -viniferin (89 mg) were obtained from fraction C-D₁₂₋₁₄ by column chromatography over Sephadex LH-20 eluted with MeOH. Gnetin E (17 mg) and latifolol (25 mg) were obtained from fraction E by column chromatography over Sephadex LH-20 eluted with MeOH followed by preparative TLC developed in benzene-AcOEt-MeOH-H₂O (20:14:6:1).

Gnetofuran A (1): white amorphous powder; $[α]^{26}_D - 7^\circ$ (*c* 0.2 MeOH); UV (MeOH) $λ_{max}$ (log ε) 308 (4.33), 288 (4.25), 220

(sh) (4.71) nm; ¹H NMR (CD₃COCD₃, 300 MHz) δ 8.06 (2H, s, OH-11,13), 7.47 (1H, s, OH-4'), 7.00 (1H, br s, H-6), 6.98 (1H, br s, H-2), 6.92 (1H, d, J = 2.1 Hz, H-2'), 6.89 (1H, d, J = 16.4 Hz, H-7), 6.78 (1H, d, J = 16.4 Hz, H-8), 6.75 (1H, dd, J = 8.1, 2.1 Hz, H-6'), 6.67 (1H, d, J = 8.1 Hz, H-5'), 6.41 (2H, d, J = 2.1 Hz, H-10,14), 6.14 (1H, t, J = 2.1 Hz, H-12), 5.45 (1H, d, J = 5.4 Hz, H-7'), 4.01 (1H, br s, OH-9'), 3.74 (2H, m, 2H-9'), 3.73 (3H, s, OCH₃-3), 3.69 (3H, s, OCH₃-3'), 3.41 (1H, dt, J= 5.4, 12.2 Hz, H-7'); ¹³C NMR (CD₃COCD₃, 75 MHz) δ 159.6 (2×s, C-11,13), 149.3 (s, C-4), 148.4 (s, C-3'), 147.3 (s, C-4'), 145.3 (s, C-3), 140.8 (s, C-9), 134.3 (s, C-1'), 132.0 (s, C-1), 130.5 (s, C-5), 129.5 (d, C-7), 127.2 (d, C-8), 119.6 (d, C-6'), 116.4 (d, C-6), 115.7 (d, C-5'), 111.9 (d, C-2), 110.5 (d, C-2'), 105.6 (2×d, C-10,14), 102.7 (d, C-12), 88.6 (d, C-7'), 64.6 (t, C-9'), 56.4 (q, OCH₃-3), 56.2 (q, OCH₃-3'), 54.7 (d, C-8'); FABMS m/z 435 [M - H]⁻; HRFABMS *m*/*z* 435.1449 (calcd for C₂₅H₂₃O₇, 435.1444).

Gnetofuran B (2): white amorphous powder; UV (MeOH) λ_{max} (log ϵ) 321 (4.03), 303 (4.21), 220 (4.50), 216 (sh) (4.59) nm; ¹H NMR (CD₃COCD₃, 400 MHz) δ 8.45 (1H, s, OH-5'), 7.46 (1H, s, OH-5), 7.35 (1H, d, J = 0.9 Hz, H-3), 7.10 (1H, dd, J = 8.8, 0.9 Hz, H-7), 7.00 (1H, t, J = 2.0 Hz, H-6'), 6.99 (1H, t, J = 2.0 Hz, H-4'), 4.05 (3H, s, OCH₃-4), 3.85 (3H, s, OCH₃-3'); ¹³C NMR (CD₃COCD₃, 100 MHz) δ 162.3 (s, C-3'), 159.8 (s, C-5'), 156.3 (s, C-2), 150.7 (s, C-7a), 144.7 (s, C-5), 140.0 (d, C-7), 105.2 (d, C-6'), 102.7 (d, C-4'), 102.5 (d, C-2'), 100.3 (d, C-3), 60.6 (q, OCH₃-4) 55.2 (q, OCH₃-3'); EIMS *mlz* 286 (M3⁺) (100), 271 (89), 243 (11), 228 (9), 149 (22), 144 (12); HREIMS *mlz* 286.0834 (calcd for C₁₆H₁₄O₅, 286.0841).

Gnetofuran C (3): white amorphous powder; UV (MeOH) λ_{max} (log ϵ) 322 (4.28), 304 (4.43), 221 (4.63) nm; ¹H NMR (CD₃-COCD₃, 400 MHz) δ 8.43 (2H, s, OH-3',5'), 7.55 (1H, s, OH-5), 7.26 (1H, br s, H-3), 7.08 (1H, br d, J = 8.0 Hz, H-7), 6.91 (2H, br s, H-2',6'), 6.85 (1H, d, J = 8.0 Hz, H-6), 6.04 (1H, br s, H-4'), 4.05 (3H, s, OCH₃-4); ¹³C NMR (CD₃COCD₃, 100 MHz) δ 159.8 (2×s, C-3',5'), 156.5 (s, C-2), 150.7 (s, C-7a), 144.8 (s, C-5), 140.0 (s, C-4), 133.0 (s, C-1'), 122.4 (s, C-3a), 114.4 (d, C-6), 106.3 (d, C-7), 104.3 (2×d, C-2',6'), 104.0 (d, C-4'), 60.6 (q, OCH₃-4); EIMS *m*/*z* 272 [M]⁺ (92), 257 (100), 228 (12), 185 (15), 149 (20), 101 (10), 59 (20); HREIMS *m*/*z* 272.0677 (calcd for C₁₅H₁₂O₅, 272.0685).

Dihydropinosylvindiol (4): white amorphous powder; $[\alpha]^{26}_{D} - 27^{\circ}$ (*c* 0.2 MeOH); UV (MeOH) λ_{max} (log ϵ) 283 (3.55), 279 (3.51), 228 (sh) (4.36) nm; ¹H NMR (CD₃COCD₃, 400 MHz) δ 7.97 (2H, s, O*H*-11,13), 7.28 (2H, br d, J = 8.0 Hz, H-2,6), 7.19 (3H, m, H-3,4,5), 6.30 (2H, d, J = 2.4 Hz, H-10,14), 6.20 (1H, t, J = 2.4 Hz, H-12), 4.72 (1H, dd, J = 6.0, 4.4 Hz, H-7), 4.63 (1H, dd, J = 6.0, 4.4 Hz, H-8), 4.08 (1H, d, J = 4.4 Hz, O*H*-7), 4.05 (1H, d, J = 4.4 Hz, O*H*-8); ¹³C NMR (CD₃COCD₃, 100 MHz) δ 158.7 (2×s, C-11,13), 145.5 (s, C-9), 143.1 (s, C-1), 128.3 (2×d, C-2,6), 128.1 (2×d, C-3,5), 127.6 (d, C-4), 106.7 (2×d, C-10,14), 102.2 (d, C-12), 78.6 (d, C-8), 78.5 (d, C-7); EIMS m/z 228 [M - H₂O]⁺ (17), 212 (11), 199 (26), 181 (12), 140 (100), 111 (23), 107 (18), 77 (16); FABMS m/z 245 [M - H]⁻; HRFABMS m/z 245.0808 (calcd for C₁₄H₁₃O₄, 245.0814).

Gnetofolin F (5): white amorphous powder; $[\alpha]^{26} D 0^{\circ} (c 0.5)$ MeOH); UV (MeOH) λ_{max} (log ϵ) 282 (4.24), 230 (4.62) nm; ¹H NMR (CD₃COCD₃, 400 MHz) & 8.13 (1H, s, OH-13), 7.81 (1H, s, OH-11), 7.48 (1H, s, OH-4), 7.23 (1H, s, OH-4'), 7.03 (1H, d, *J* = 2.0 Hz H-2), 6.89 (1H, dd, *J* = 8.0, 2.0 Hz, H-6), 6.82 (1H, d, J = 8.0 Hz, H-5), 6.73 (1H, d, J = 2.0 Hz, H-2'), 6.66 (1H, d, J = 8.0 Hz, H-5'), 6.50 (1H, dd, J = 8.0, 2.0 Hz, H-6'), 6.35 (1H, d, J = 2.0 Hz, H-14), 6.27 (1H, d, J = 2.0 Hz, H-12), 4.71 (1H, d, J = 4.0 Hz, H-7), 4.46 (1H, t, J = 8.8 Hz, H-9'_{β}), 4.18 (1H, s, H-7'), 3.81 (1H, dd, J = 8.8, 4.0 Hz, H-8), 3.51 (1H, t, t)J = 8.8 Hz, H-9'_a), 3.03 (1H, q, J = 8.8 Hz, H-8), 3.86 (3H, s, OCH₃-3), 3.74 (3H, s, OCH₃-3'); ¹³C NMR (CD₃COCD₃, 100 MHz) δ 159.8 (s, C-13), 155.8 (s, C-11), 148.3 (2×s, C-3,9), 148. 0 (s, C-3'), 146.7 (s, C-4), 145.5 (s, C-4'), 138.0 (s, C-1'), 135.5 (s, C-1), 122.8 (s, C-10), 120.3 (d, C-6'), 119.6 (d, C-6), 115.5 (d, C-5), 115.4 (d, C-5'), 111.9 (d, C-2'), 110.5 (d, C-2), 103.3 (d, C-14), 102.7 (d, C-12), 88.3 (d, C-7), 74.5 (t, C-9'), 59.7 (d, C-8), 56.2 (q, OCH3-3), 56.1 (q, OCH3-3'), 55.9 (d, C-8'), 51.0 (d, C-7'); FABMS m/z 435 [M - H]-; HRFABMS m/z 435.1448 (calcd for C₂₅H₂₃O₇, 435.1444).

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NP020532O